

been used. This is adequate to keep molds from appearing and seems not to give bad effects on the flies.

Bridges, C. B. On the seeding of culture with yeast.

On some five occasions the method of seeding the surfaces of culture bottles by spraying with very thin yeast suspension (see DIS-6:66) has been hopefully put into practice and then abandoned

in favor of seeding with one, or better two fat drops of thick yeast suspension. Always the failures of pair cultures to produce offspring became so large as to seriously hinder the experimental breeding. The main advantage of the spraying was the suppression of mold growth - and this is now better accomplished by moldex.

Bridges, C. B. On distinguishing larvae for salivary preparations.

The notes by Beadle (Am. Nat. 71: 277; DIS-6:24), Hoover (DIS-6:24) and Brehme (DIS-8: - ) show how the distinction between the yellow of normal malpighian tubules and the colorless or paler tubules

associated with certain light eye-colors (notably w and lt but also cm,  $g^2$   $p^D$  and ca) and the brown color of the mouth parts of yellow larvae, can be used to select larvae of the type desired for salivary preparations. A survey of our balancers shows that a few of them are especially useful for general use in balancing any mutant whose salivaries might need investigation. For chromosome I, these are: Cl, y Hw (carrying y and  $g^2$ ) dl-49,  $m^2$   $g^4$  (carrying  $g^4$ , already recommended by Hoover) and dl-49, w lz<sup>s</sup>. For chromosome II, the best is Cy, al<sup>2</sup> lt<sup>3</sup> L<sup>4</sup> sp<sup>2</sup> (carrying lt<sup>3</sup> of Beadle) and for chromosome III, the best is Payne, Dfd ca. These balancers are the best of the ClB, dl-49, Cy and Payne varieties, and should be kept on hand and favored in stock making.

For second and third chromosome aberrations, use can be made of the dominant eye-color  $p^G$  which has pale tubules (perhaps  $p^G/p^D$  are better). First cross to T(Y;2;3) $p^G$ , pick out the T(2;3) $p^G$  aberration  $\sigma^D$  and cross to  $p^D$   $\phi$ , using the normal yellow tubuled larvae.

A second and third chromosome method needing no distinguishing of larvae, except the easy one of femaleness, has been the use (by Schultz and myself) of T(Y;2;3)I. Cross the female bearing the aberration to  $\phi$  males carrying T(Y;2;3)I, pick out sons carrying the aberration (all are T(Y;2;3)I) and cross to any standard female. All daughters are heterozygotes for the aberration.

Buzzati-Traverso, A. Method for making salivary gland chromosomes permanent smearing.

I found very convenient for making permanent salivary gland chromosome smears the following method: (1) Dissect as usual the larva and leave the salivary gland in normal aceto carmine till well stained; (2) pass the gland

to the slide and take off all the aceto carmine which might